ab108787 – Albumin Human ELISA Kit

Instructions for Use

For the quantitative measurement of Human Albumin in plasma and serum.

Version 9b Last Updated 12 December 2023

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1. <u>BACKGROUND</u>

Abcam's Albumin Human *in vitro* competitive ELISA (Enzyme-Linked Immunosorbent Assay) kit is designed for the measurement of albumin in Human plasma and serum.

An albumin specific antibody has been precoated onto 96-well plates and blocked. Standards or test samples are added to the wells and subsequently biotinylated albumin is added and then followed by washing with wash buffer. Streptavidin-Peroxidase Complex is added and unbound conjugates are washed away with wash buffer. TMB is then used to visualize Streptavidin-Peroxidase enzymatic reaction. TMB is catalyzed by Streptavidin-Peroxidase to produce a blue color product that changes into yellow after adding acidic stop solution. The density of yellow coloration is inversely proportional to the amount of albumin captured in plate.

Albumin is serum hepatic protein, the most abundant protein in serum and contributes to the maintenance of oncotic pressure as well as to the transport of hydrophobic molecules. Serum albumin level has been linked in clinical practice to several diseases. Low albumin levels can suggest liver disease, kidney disease, inflammation, shock, and malnutrition. On the other hand, high albumin levels usually reflect dehydration.

2. <u>ASSAY SUMMARY</u>

Primary Capture Antibody
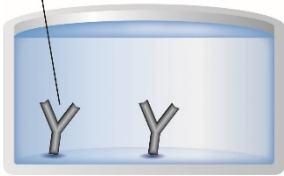
Prepare all reagents, samples and standards as instructed.

Sample   Biotinylated Antigen
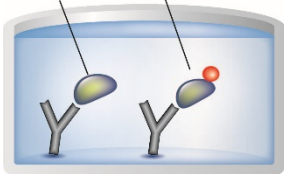
Add standard or sample to each well used and add prepared biotin protein to each well. Incubate at room temperature.

Strepavidin-HRP
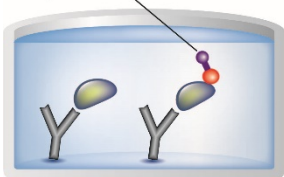
Wash and add prepared Streptavidin-Peroxidase Conjugate. Incubate at room temperature.

Substrate   Colored Product

Wash and add Chromogen Substrate to each well. Incubate at room temperature. Add Stop Solution to each well. Read immediately.

3. PRECAUTIONS

Please read these instructions carefully prior to beginning the assay.

Modifications to the kit components or procedures may result in loss of performance.

4. STORAGE AND STABILITY

Store kit at 4°C immediately upon receipt, apart from the SP Conjugate & Biotinylated Protein, which should be stored at -20°C.
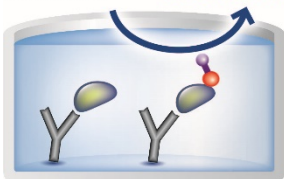
Refer to list of materials supplied for storage conditions of individual components. Observe the storage conditions for individual prepared components in sections 9 & 10.

5. MATERIALS SUPPLIED

Item	Amount	Storage Condition (Before Preparation)
Albumin Microplate (12 x 8 well strips)	96 wells	4°C
Albumin Standard	1 vial	4°C
10X Diluent N Concentrate	30 mL	4°C
Biotinylated Human Albumin (Lyophilized)	1 vial	-20°C
100X Streptavidin-Peroxidase Conjugate (SP Conjugate)	80 µL	-20°C
Chromogen Substrate	7 mL	4°C
Stop Solution	11 mL	4°C
20X Wash Buffer Concentrate	30 mL	4°C
Sealing Tapes	3	N/A

6. MATERIALS REQUIRED, NOT SUPPLIED

These materials are not included in the kit, but will be required to successfully utilize this assay:

- 1 Microplate reader capable of measuring absorbance at 450 nm.
- Precision pipettes to deliver 1 µL to 1 mL volumes.
- Adjustable 1-25 mL pipettes for reagent preparation.
- 100 mL and 1 liter graduated cylinders.
- Absorbent paper.
- Distilled or deionized water.
- Log-log graph paper or computer and software for ELISA data analysis.
- 5 tubes to prepare standard or sample dilutions.

7. LIMITATIONS

- Do not mix or substitute reagents or materials from other kit lots or vendors.

8. TECHNICAL HINTS

- Samples generating values higher than the highest standard should be further diluted in the appropriate sample dilution buffers.

- Avoid foaming or bubbles when mixing or reconstituting components.

- Avoid cross contamination of samples or reagents by changing tips between sample, standard and reagent additions.

- Ensure plates are properly sealed or covered during incubation steps.

- Complete removal of all solutions and buffers during wash steps.

- **This kit is sold based on number of tests. A 'test' simply refers to a single assay well. The number of wells that contain sample, control or standard will vary by product. Review the protocol completely to confirm this kit meets your requirements. Please contact our Technical Support staff with any questions.**

9. REAGENT PREPARATION

Equilibrate all reagents to room temperature (18-25°C) prior to use. Prepare fresh reagents immediately prior to use. If crystals have formed in the concentrate, mix gently until the crystals have completely dissolved.

9.1 **1X Diluent N**

Dilute the 10X Diluent N Concentrate 1:10 with reagent grade water to produce a 1x solution. Mix gently and thoroughly. *Store for up to 30 days at 4°C.*

9.2 **1X Wash Buffer**

Dilute the 20X Wash Buffer Concentrate 1:20 with reagent grade water to produce a 1x solution. Mix gently and thoroughly.

9.3 **1X Biotinylated Albumin**

Add 4 mL 1X Diluent N to the lyophilized Biotinylated Albumin vial to generate a 2X Biotinylated Albumin stock solution. Allow the vial of 2X Biotinylated Albumin stock solution to sit for 10 minutes with gentle agitation prior to use.

The stock solution should be further diluted 1:2 with 1X Diluent N to generate 1X Biotinylated Albumin.

Any remaining solution should be frozen at -20°C.

9.4 **1X SP Conjugate**

Spin down the 100X Streptavidin-Peroxidase Conjugate (SP Conjugate) briefly and dilute the desired amount of the conjugate 1:100 with 1X Diluent N, to produce a 1x solution.

Any remaining solution should be frozen at -20°C.

10. STANDARD PREPARATIONS

- **Prepare serially diluted standards immediately prior to use. Always prepare a fresh set of standards for every use.**

- **Any remaining standard should be stored at -20°C after reconstitution and used within 30 days.**

- **This procedure prepares sufficient standard dilutions for duplicate wells.**

  10.1 Reconstitution of the Albumin Standard vial to prepare the 100 µg/mL Albumin **Standard #1**:

      10.1.1 First consult the Albumin Standard vial to determine the mass of protein in the vial.

      10.1.2 Calculate the appropriate volume of 1X Diluent N to add when resuspending the Albumin Standard vial to produce a 100 µg/mL Albumin **Standard #1** by using the following equation:

  C_S = Starting mass of Albumin Standard (see vial label) (µg)

  C_F = The 100 µg/mL Albumin **Standard #1** final required concentration

  V_D = Required volume of 1X Diluent N for reconstitution (µL)

  Calculate total required volume 1X Diluent N for resuspension:

  $$(C_S / C_F) \times 1{,}000 = V_D$$

  Example:

  NOTE: This example is for demonstration purposes only. Please remember to check your standard vial for the actual amount of standard provided.

  C_S = 130 µg of Albumin Standard in vial

  C_F = 100 µg/mL Albumin **Standard #1** final concentration

 V_D = Required volume of 1X Diluent N for reconstitution

  $$(130 \text{ µg} / 100 \text{ µg/mL}) \times 1{,}000 = 1{,}300 \text{ µL}$$

10.1.3 Reconstitute the Albumin Standard vial by adding the appropriate calculated amount V_D of 1X Diluent N to the vial to generate the 100 µg/mL **Standard #1**. Mix gently and thoroughly.

10.2 Allow the reconstituted 100 µg/mL **Standard #1** to sit for 10 minutes with gentle agitation prior to making subsequent dilutions

10.3 Label five tubes #2 – 6

10.4 Add 360 µL of 1X Diluent N to tube #2 – 6.

10.5 To prepare **Standard #2**, add 120 µL of the **Standard #1** into tube #2 and mix gently.

10.6 To prepare **Standard #3**, add 120 µL of the **Standard #2** into tube #3 and mix gently.

10.7 Using the table below as a guide, prepare subsequent serial dilutions.

10.8 1X Diluent N serves as the zero standard, 0 µg/mL (tube #6).

Standard Dilution Preparation Table

Standard #	Volume to Dilute (µL)	Volume Diluent N (µL)	Total Volume (µL)	Starting Conc. (µg/mL)	Final Conc. (µg/mL)
1	Step 10.1				100.0
2	120	360	480	100.0	25.00
3	120	360	480	25.00	6.250
4	120	360	480	6.250	1.563
5	120	360	480	1.563	0.391
6	-	360	360	-	0
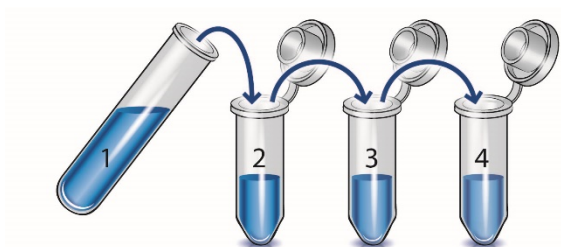
11. SAMPLE PREPARATION

11.1 Plasma

Collect plasma using one-tenth volume of 0.1 M sodium citrate as an anticoagulant. Centrifuge samples at 3,000 x *g* for 10 minutes. Dilute samples 1:10,000 into 1X Diluent N and assay. The undiluted samples can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles. (EDTA or Heparin can also be used as an anticoagulant.)

11.2 Serum

Samples should be collected into a serum separator tube. After clot formation, centrifuge samples at 3,000 x *g* for 10 minutes and remove serum. Dilute samples 1:10,000 into 1X Diluent N and assay. The undiluted serum can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.

12. PLATE PREPARATION

- The 96 well plate strips included with this kit are supplied ready to use. It is not necessary to rinse the plate prior to adding reagents.
- Unused well plate strips should be returned to the plate packet and stored at 4°C.
- For statistical reasons, we recommend each sample should be assayed with a minimum of two replicates (duplicates).
- Well effects have not been observed with this assay. Contents of each well can be recorded on the template sheet included in the Resources section.

13. <u>ASSAY PROCEDURE</u>

- **Equilibrate all materials and prepared reagents to room temperature (18-25°C) prior to use.**

- **It is recommended to assay all standards, controls and samples in duplicate.**

  13.1   Prepare all reagents, standard solutions and samples as instructed. Equilibrate reagents to room temperature before use. The assay is performed at room temperature (18-25°C).

  13.2   Remove excess microplate strips from the plate frame and return them immediately to the foil pouch with desiccant inside. Reseal the pouch securely to minimize exposure to water vapor and store in a vacuum desiccator.

  13.3   Add 25 µL of Albumin Standard or sample per well and immediately add 25 µL of 1X Biotinylated Albumin to each well (on top of Standard or sample). Cover wells with a sealing tape and incubate for one hour at room temperature. Start the timer after the last addition.

  13.4   Wash five times with 200 µL of 1X Wash Buffer manually. Invert the plate each time and decant the contents; tap it 4-5 times on absorbent paper towel to completely remove the liquid. If using a machine wash six times with 300 µL of 1X Wash Buffer and then invert the plate, decant the contents; tap it 4-5 times on absorbent paper towel to completely remove the liquid.

  13.5   Add 50 µL of 1X SP Conjugate to each well, cover with a sealing tape and incubate for 30 minutes. Turn on the microplate reader and set up the program in advance.

  13.6   Wash microplate as described above.

  13.7   Add 50 µL of Chromogen Substrate per well and incubate in ambient light for 20 minutes or until the optimal blue colour density develops. Gently tap plate to ensure

thorough mixing and break the bubbles in the well with pipette tip.

13.8   Add 50 µL of Stop Solution to each well. The color will change from blue to yellow.

13.9   Read the absorbance on a microplate reader at a wavelength of 450 nm <u>immediately</u>. If wavelength correction is available, subtract readings at 570 nm from those at 450 nm to correct optical imperfections. Otherwise, read the plate at 450 nm only. Please note that some unstable black particles may be generated at low concentration points after stopping the reaction for about 10 minutes, which will reduce the readings.

14. CALCULATIONS

Calculate the mean value of the triplicate readings for each standard and sample. To generate a Standard Curve, plot the graph using the standard concentrations on the x-axis and the corresponding mean 450 nm absorbance on the y-axis. The best-fit line can be determined by regression analysis using log-log or four-parameter logistic curve-fit. Determine the unknown sample concentration from the Standard Curve and multiply the value by the dilution factor.

15. TYPICAL DATA

TYPICAL STANDARD CURVE – Data provided for demonstration purposes only. A new standard curve must be generated for each assay performed.
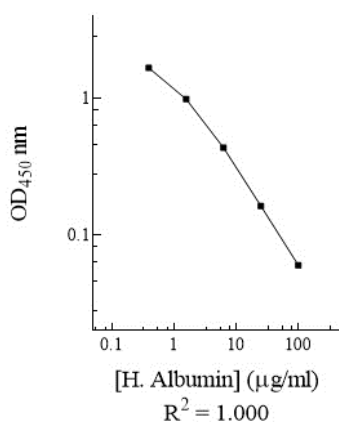
Human Albumin Standard Curve

Figure 1. Example of a Human Albumin standard curve prepared as described in Section 10.

16. TYPICAL SAMPLE VALUES

SENSITIVITY –

The minimum detectable dose of Human Albumin is typically ~0.28 µg/mL.

RECOVERY –

Standard Added Value: 1.5 – 25 µg/mL

Recovery %: 91 – 110.

Average Recovery %: 98

REFERENCE VALUE –

The normal Human plasma levels of Albumin are 34-54 mg/mL.

LINEARITY OF DILUTION –

Plasma Dilution	Average % Expected Value
1:5,000	104
1:10,000	101
1:20,000	95

Serum Dilution	Average % Expected Value
1:5,000	102
1:10,000	99
1:20,000	97

PRECISION –

	Intra-Assay	Inter-Assay
% CV	3.3	10.6

17. ASSAY SPECIFICITY

Species	% Cross Reactivity
Canine	None
Mouse	None
Monkey	None
Bovine	None
Rat	None
Swine	None
Rabbit	None
Equine	None

18. TROUBLESHOOTING

Problem	Cause	Solution
Poor standard curve	Improper standard dilution	Confirm dilutions made correctly
	Standard improperly reconstituted (if applicable)	Briefly spin vial before opening; thoroughly resuspend powder (if applicable)
	Standard degraded	Store sample as recommended
	Curve doesn't fit scale	Try plotting using different scale
Low signal	Incubation time too short	Try overnight incubation at 4°C
	Target present below detection limits of assay	Decrease dilution factor; concentrate samples
	Precipitate can form in wells upon substrate addition when concentration of target is too high	Increase dilution factor of sample
	Using incompatible sample type (e.g. serum vs. cell extract)	Detection may be reduced or absent in untested sample types
	Sample prepared incorrectly	Ensure proper sample preparation/dilution
Large CV	Bubbles in wells	Ensure no bubbles present prior to reading plate
	All wells not washed equally/thoroughly	Check that all ports of plate washer are unobstructed wash wells as recommended
	Incomplete reagent mixing	Ensure all reagents/master mixes are mixed thoroughly
	Inconsistent pipetting	Use calibrated pipettes and ensure accurate pipetting
	Inconsistent sample preparation or storage	Ensure consistent sample preparation and optimal sample storage conditions (eg. minimize freeze/thaws cycles)

Problem	Cause	Solution
High background/ Low sensitivity	Wells are insufficiently washed	Wash wells as per protocol recommendations
	Contaminated wash buffer	Make fresh wash buffer
	Waiting too long to read plate after adding STOP solution	Read plate immediately after adding STOP solution
	Improper storage of ELISA kit	Store all reagents as recommended. Please note all reagents may not have identical storage requirements.
	Using incompatible sample type (e.g. Serum vs. cell extract)	Detection may be reduced or absent in untested sample types

19. NOTES

Technical Support

For all technical or commercial enquiries please go to:
www.abcam.com/contactus
www.abcam.cn/contactus (China)
www.abcam.co.jp/contactus (Japan)